

Tuning the Rate of Molecular Translocation

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Biological molecules contain dynamic and functional interiors capable of directing chemical operations such as gating, translational/rotational movement, and covalent bond formation/cleavage.¹ These natural systems have their working components synchronized² and well-ordered by the hierarchical constitution.³ Abiotic molecules/assemblies, adept at mediating encapsulation or chemical reactions,⁴ are less sophisticated but nonetheless designed to operate with intricate mechanisms.⁵ Self-folding cavitands⁶ have, for instance, been made to entrap/release molecules via folding, and (hemi)carcerands are known to prolong the lifetime and modulate the reactivity of “fleeting” intermediates.⁷ Transition metal assembled cages, additionally, facilitate chemical transformations by virtue of their discrete inner-space characteristics.⁸ Alongside these accomplishments in the field, gating has been recognized⁹ as an important element affecting the kinetic lability (dissociation)¹⁰ of encapsulation complexes. Full control of the gating presents an opportunity for controlling the kinetic reactivity in abiotic hosts; however, this control still poses a challenge.¹¹ This study, consequently, focuses on examining the in/out rate of guest’s encapsulation as a function of the dynamics of gates revolving at the rim of gated molecular baskets (Figure 1).¹² Quantitative relationships have been established to allow for predicting and controlling the time that guest molecules spend inside the basket’s interior.

Molecular baskets have been designed^{12a} to contain three pyridine-based gates, linked via intramolecular hydrogen bonding (HB) from *meta* amido groups, to occlude space and thus form a dynamic and gated environment (Figure 1). The “hinge” H_{a/b} signals appeared as a singlet at high temperatures and as an AB quartet at low temperatures, thus demonstrating the interconversion of two C₃ symmetric enantiomers, **A** and **B**, each containing hydrogen bonds displayed in a clockwise or counterclockwise orientation (Figure 1B).^{12,14} In fact, the **A/B** interconversion necessitates that each of the gates revolves 180° about the “vertical” axis. This conformational change has additionally been shown to correlate with the exchange (departure/entrance) of guest molecules.^{12a} To quantitatively examine the structure–activity relationship¹³ in this supramolecular environment, we chose to alter the electronic (inductive and field) and steric (bulkiness) characteristics of the R amido units (see **1–6**, Figure 1). The hypothesis was that the perturbation imposed by R groups would, in **1–6**, affect the host’s conformational dynamics and thereby allow a precise and predictable fine-tuning of the guest’s kinetic lability.

Six R groups were deliberately chosen and installed to give baskets **1–6** (Figure 1A). The syntheses proceeded via methodology whereby *tris*-anhydride **7** was reacted with 5-(aminomethyl)pyridin-3-amine to yield modular *tris*-amine **8**.¹⁴ Subsequently, an alkanoylation of **8** gave desired **2–6** in satisfactory yields (70–85%).

The electron density perturbation in **1–6**, caused by substituents R, ought to anisotropically impact the electrostatic N–H···N contacts and thereby have an effect on the intramolecular hydrogen

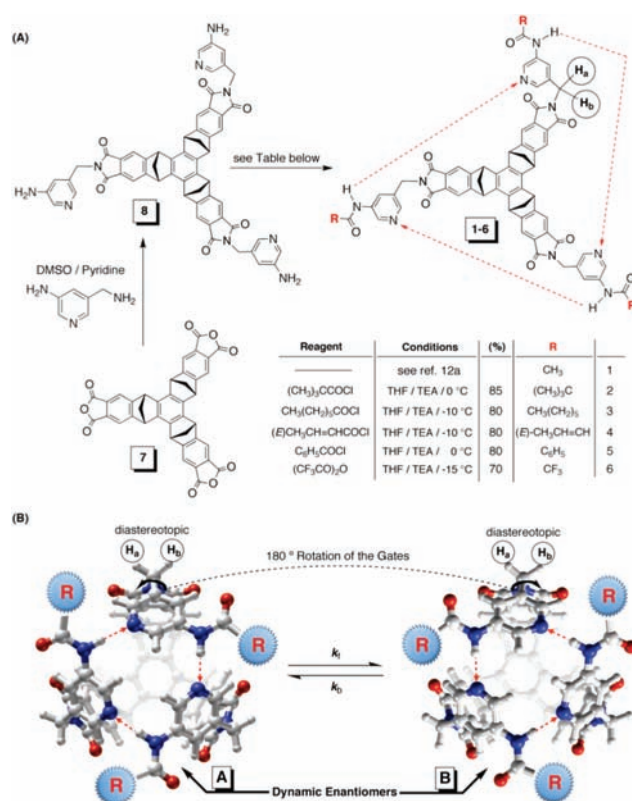


Figure 1. Chemical structures of molecular baskets **1–6** (A). Top view of **A** and **B** dynamic enantiomers of **1–6** that interconvert by simultaneous 180° rotation of the gates (B). The diastereotopic nature of H_{a/b} protons permitted a thermal dependence of the ¹H NMR line shapes, which upon simulation analysis gave the interconversion rate constants (k_{1/2}).¹⁴

Table 1. Calculated Electrostatic Potential Energies (AM1/HF (6-31G**)) at the N–H and Pyr–N Sites in Model Compounds¹⁴ and ¹H NMR (400 MHz, CD₂Cl₂) Chemical Shifts of the N–H Resonance of **1–6** Containing an Excess (> 60 mol equiv) of *t*-BuBr

| R | N–H ^a (kcal/mol) | Pyr–N ^a (kcal/mol) | δ (N–H) ^b (ppm) | δ (N–H) ^c (ppm) |
|---|--------------------------------|----------------------------------|-------------------------------|-------------------------------|
| (CH ₃) ₃ C | 58 | –49 | 9.7 | 10.4 |
| CH ₃ | 62 | –48 | 10.8 | 11.4 |
| CH ₃ CH=CH | 61 | –49 | 10.8 | 11.4 |
| CH ₃ (CH ₂) ₅ | 62 | –49 | 10.7 | 11.3 |
| C ₆ H ₅ | 59 | –49 | 11.2 | 11.8 |
| CF ₃ | 69 | –44 | 12.2 | 12.6 |

^a Model compounds¹⁴ consist of one phthalimide “arm”. ^b 298 K. ^c 213 K.

bonding.¹⁵ That is to say, the depletion (or build-up) of the charge at the HB-donor position (N–H) must be accompanied by a negligible charge perturbation at the HB-acceptor site (Pyr–N). Indeed, the electrostatic potentials of the energy minimized model

Table 2. Kinetic Parameters for the Revolving of Gates (k_b , ^1H NMR Line-Shape Analysis) and the Translocation of $t\text{-BuBr}$ (k_{in} , k_{out} , 2D EXSY NMR) in Molecular Baskets **1–6** (CD_2Cl_2), at 226.0 ± 0.1 K. Thermodynamic Stabilities (ΔG° , 226.0 K) of the Encapsulation Complexes

| basket | R | k_b (s^{-1}) ^{a,b} | k_{in} ($\text{M}^{-1} \text{s}^{-1}$) ^b | k_{out} (s^{-1}) ^b | ΔG_b^\ddagger (kcal/mol) | ΔG_{out}^\ddagger (kcal/mol) | ΔG° (kcal/mol) |
|----------|--------------------------------------|--|---|--|----------------------------------|--------------------------------------|-----------------------------|
| 1 | CH_3 | 108 ± 22 | 524 ± 110 | 4.7 ± 0.7 | 11.0 ± 0.1 | 12.4 ± 0.1 | -2.1 ± 0.1 |
| 2 | $(\text{CH}_3)_3\text{C}$ | 78 ± 16 | 1964 ± 392 | 11.5 ± 0.9 | 11.1 ± 0.1 | 12.0 ± 0.1 | -2.3 ± 0.1 |
| 3 | $\text{CH}_3(\text{CH}_2)_5$ | 97 ± 20 | 407 ± 73 | 4.3 ± 0.4 | 11.0 ± 0.1 | 12.4 ± 0.1 | -2.0 ± 0.1 |
| 4 | $(E)\text{-CH}_3\text{CH}=\text{CH}$ | 83 ± 17 | 239 ± 68 | 2.8 ± 0.1 | 11.1 ± 0.1 | 12.6 ± 0.1 | -2.0 ± 0.1 |
| 5 | C_6H_5 | 20 ± 4 | 38 ± 12 | 0.4 ± 0.1 | 11.7 ± 0.1 | 13.5 ± 0.1 | -2.0 ± 0.2 |
| 6 | CF_3 | 4 ± 1 | 0.7 ± 0.1 | 0.07 ± 0.02 | 12.4 ± 0.1 | 14.2 ± 0.2 | -1.0 ± 0.2 |

^a Error margins (20%) were obtained on the basis of four independent measurements.¹⁴ ^b Each measurement was repeated twice, and the error margins were propagated from the linear least-squares analysis of the experimental data.¹⁴

compounds¹⁴ (AM1/HF(6-31G**), Spartan)¹⁶ suggested a fluctuation in the charge density at the donor but rather consistent values at the acceptor atom (Table 1).

^1H NMR chemical shifts of the N–H signals in **1–6** are indicators for the strength and the proportion of the intramolecular hydrogen bonding (Table 1).¹⁷ Markedly, bulky $(\text{CH}_3)_3\text{C}$ – groups enforced weaker ($\delta_{\text{N-H}} = 9.7\text{--}10.4$ ppm) whereas all other substituents stronger ($\delta_{\text{N-H}} = 10.8\text{--}12.6$ ppm) noncovalent contacts. FT-IR spectroscopic studies of **1**, **2**, and **6**, in addition, suggested the existence of a fully closed C_3 symmetric basket (as shown in Figure 1B) with other conformers populating the equilibrium to a lesser degree.¹⁴ These findings are, importantly, supported by Schneider's suggestion¹⁸ that ~ 2 kcal/mol of the free energy (ΔG° , 298 K, CDCl_3) is to be attributed to a hydrogen bond lacking secondary electrostatic interactions; that is to say, ΔG° (298 K, CDCl_3) of ~ 6 kcal/mol can be expected to describe the formation of a “fully closed” basket.

First-order rate constants (k_{pb}) for the interconversion of dynamic enantiomers **A** and **B** in **1–6** (Figure 1B) were determined by completing the line-shape analysis of the diastereotopic H_{ab} resonances at variable temperatures.¹⁴ Importantly, an excess of $t\text{-BuBr}$ (guest) was used in each experiment to ensure a sole exchange of the guest-populated baskets. The rate constants (k_b) for “averaging” the H_{ab} signals, i.e., revolving of the gates, were for **1–6** further incorporated into an Eyring plot to afford k_b 's at 226.0 K (Table 2).¹⁴ It is obvious that substituents had an effect on the gates' dynamics: the electron-withdrawing CF_3 retarded ($4 \pm 0.4 \text{ s}^{-1}$) while the electron-donating CH_3 ($108 \pm 22 \text{ s}^{-1}$) accelerated the rotation.

The dependence of the reaction's free energy (ΔG° or ΔG^\ddagger) on the reactant's substituents is described with substituent constants (σ) and accounted for by proportional free energy relationships (LFERs).¹³ The concept has, interestingly, been used for examining noncovalent interactions, and typically, the electronic effects are solely evaluated.¹⁹ In the case of baskets **1–6**, however, the gates' dynamics seems to be a function of not only electronic but also steric factors (Table 2). Moreover, the ground and the excited states for the **A/B** interconversion appear crowded when sizable R groups are introduced (Figure 1B). Taft's LFER scale, conveniently, defines polar (σ^*) and steric (E_s) substituent constants and has been recommended for studying aliphatic systems.²⁰ We used this two-parameter model to fit a plot of $\log(k_b^{(\text{subs.})}/k_b^{(\text{Me})})$ versus $\rho^*\sigma^* + \delta E_s$ (Figure 2A). The correlation was acceptable ($R^2 = 0.94$), with the revolving rates susceptible to both inductive/field ($\rho^* = -0.5$) and steric ($\delta = 0.13$) factors. Molecular basket **2** with the sterically demanding $(\text{CH}_3)_3\text{C}$ – group, thus, underwent a rather “slow” **A/B** interconversion ($k_b = 78 \pm 16 \text{ s}^{-1}$) despite its weak N–H \cdots N hydrogen bonding contacts (Table 1).

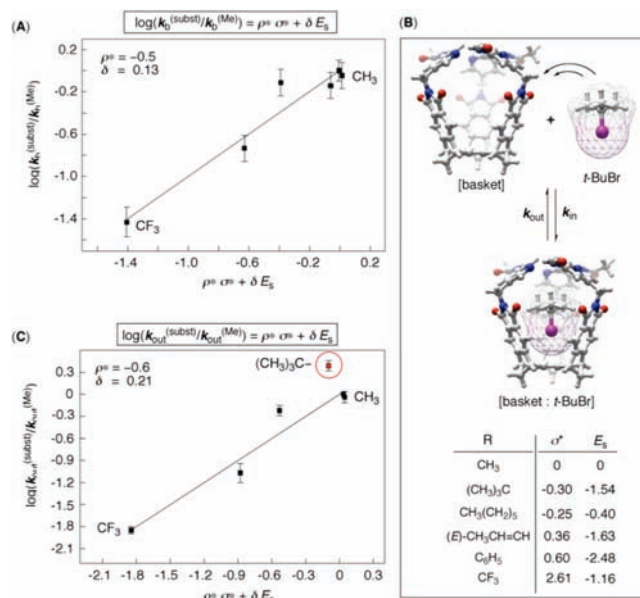


Figure 2. Linear free energy relationships (LFERs) for the revolving of gates (A) in baskets **1–6** and the dissociation of $t\text{-BuBr}$ (B/C) from baskets **1–6**. Both correlations were obtained using Taft's two-parameter regression model with polar (σ^*) and steric (E_s) substituent constants (B).

The rate constants for $t\text{-BuBr}$ (guest) entering (k_{in}) and departing (k_{out}) baskets **1–6** (Figure 2B) were ascertained by completing quantitative ^1H – ^1H NOESY NMR (exchange spectroscopy, EXSY) measurements.²¹ The volumes of the cross and diagonal peaks for proton resonances of $t\text{-BuBr}$ inside and outside the basket were evaluated to give first-order magnetization rate constants (k_{in}^* and k_{out}^*) for the exchange, at different mixing times (τ_m); τ_m 's were originally estimated by measuring T_1 relaxations of $t\text{-BuBr}$ protons.^{10b,21,22} The association k_{in} and dissociation k_{out} rate constants (Figure 2B) were then obtained as $k_{in} = k_{in}^*/[\text{basket}]$ and $k_{out} = k_{out}^*$ (Table 2).

Taft's linear free energy scale was used to correlate the kinetic data for the departure (k_{out}) of $t\text{-BuBr}$ (Figure 2C). A plot of $\log(k_{out}^{(\text{subs.})}/k_{out}^{(\text{Me})})$ versus $\rho^*\sigma^* + \delta E_s$ correlates well ($R^2 = 0.95$). The polar ($\rho^* = -0.6$) and steric ($\delta = 0.21$) sensitivity factors characterizing the rate of $t\text{-BuBr}$ dissociating from baskets **1–6** are similar to the factors describing the rotation of the gates ($\rho^* = -0.5$ and $\delta = 0.13$). The result, importantly, validates the interdependence between the internal dynamics of the gates (rotary motion) and the kinetic stability of guests (translation). Thus far, the electron-withdrawing CF_3 retarded ($k_{out} = 0.07 \pm 0.02 \text{ s}^{-1}$) while the electron-donating CH_3 ($k_{out} = 4.7 \pm 0.7 \text{ s}^{-1}$) accelerated the departure of $t\text{-BuBr}$ from the baskets. The behavior of basket **2** containing bulky $(\text{CH}_3)_3\text{C}$ – units, however, did not follow the linear trend described in Figure 2C. The guest ($t\text{-BuBr}$) departed this

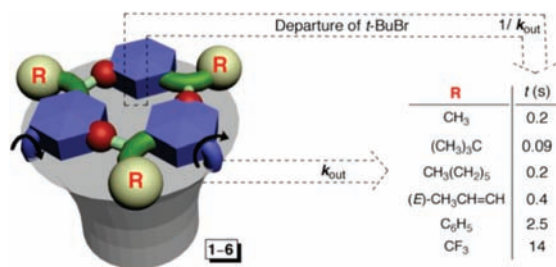


Figure 3. Schematic representation of gated molecular baskets **1–6** capable of controlling time (t) that t -BuBr spends in their cavity (right). Choosing a proper R substituent, one can now tune this “residing” time to a desired value.

basket at a rate ($k_{\text{out}} = 11.5 \pm 0.9 \text{ s}^{-1}$) higher than expected considering the “slow” flipping of its gates ($k_b = 78 \pm 16 \text{ s}^{-1}$). It is, perhaps, that weaker hydrogen bonds at the seam of this basket (Table 1) authorized the guest departure by an alternative mechanism. One scenario could involve the slipping of t -BuBr through an aperture created by single-gate unfolding, but this remains to be further investigated.

The stability of t -BuBr complexes with **1–6** (ΔG° , Table 2), apparently, decreased throughout the series. A notion that a more tightly closed basket has a “smaller” inner space, and thereby higher affinity toward solvent and lower toward t -BuBr, is appealing. The observed opposition of the thermodynamic stability and kinetic lability in the guest binding could, however, be important for catalytic applications.

The lifetime t ($1/k_{\text{out}}$) of the encapsulated t -BuBr is clearly a function of the basket’s dynamics (Figure 3) and can in this gated environment be controlled and predicted by choosing a proper R substituent. Considering the potential of encapsulating and stabilizing transient species²³ inside gated molecular baskets, this method of regulating the kinetic lability will now be extended toward directing the course of chemical reactions.

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Supporting Information Available: Detailed description of experimental methods. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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